An Integrated Flow and Batch-Based Approach for the Synthesis of *O*-Methyl Siphonazole

Marcus Baumann, Ian R. Baxendale, Malte Brasholz, John J. Hayward, Steven V. Ley, Nikzad Nikbin

Innovative Technology Centre, Department of Chemistry, University of Cambridge, Lensfield Road, Cambridge CB2 1EW, UK E-mail: theitc@cam.ac.uk

Received 21 February 2011

Abstract: The bisoxazole containing natural product *O*-methyl siphonazole was assembled using a suite of microreactors via a flowbased approach in concert with traditional batch methods. The use of a toolbox of solid-supported scavengers and reagents to aid purification afforded the natural product in a total of nine steps.

Key words: oxazoles, Claisen condensation, flow chemistry, microreactors, solid-supported reagents

As part of our ongoing interest in oxazole-containing natural products,¹ we have devised a synthetic route to the bisoxazole alkaloid *O*-methyl siphonazole (**2**), which was discovered by König and co-workers in 2006, along with the C-21-demethylated parent compound siphonazole (**1**, Scheme 1).² While Moody³ and Ciufolini⁴ reported the first syntheses of the siphonazoles in 2008 and 2009, a detailed biological profile of these metabolites of *Herpetosiphon sp.* has not been thoroughly established. Thus, a modular synthesis of the natural products **1** and **2**, flexible enough for the production of unnatural analogues, is of interest.

In this article we disclose our results on the preparation of O-methyl siphonazole (2) based on an integrated approach combining conventional batch and new flow chemistry methods. The application of flow techniques in organic synthesis as described by numerous laboratories⁵

has transformed flow chemistry from an academic interest to a valuable enabling technology that overcomes several of the bottlenecks traditionally faced by synthetic chemists.⁶ As such the generation and immediate use of hazardous, toxic, or unstable intermediates⁷ has been reported as well as the possibility to more reliably perform reaction scale-up.8 Furthermore, flow reactors can be transformed into automated platforms by the addition of liquid handling and fraction collector modules expanding the working capabilities of the units and allowing for 24/7 working regimes.9 Finally, individual reactions can be telescoped into one continuous flow sequence, thus avoiding the iterative isolation, purification, and reprocessing of intermediates.¹⁰ With these potential advantages in mind, we have orchestrated an improved total synthesis of O-methyl siphonazole by making use of flow techniques in order to expedite certain steps in the synthesis. Our first synthetic target was the oxazole carboxylic acid 10 which we intended to access from commercially available dimethoxycinnamic acid (6, Scheme 2).

In order to process this material quickly we decided to employ the Vapourtec R2+/R4 flow system¹¹ where two small HPLC pumps deliver streams of starting materials which are subsequently mixed at either a T-piece or within a microchip. The resulting stream can then be introduced into a tubular convection flow coil (CFC) which is



Scheme 1 Siphonazole (1) and its building blocks 3–5

SYNLETT 2011, No. 10, pp 1375–1380 Advanced online publication: 23.05.2011 DOI: 10.1055/s-0030-1260573; Art ID: D05911ST © Georg Thieme Verlag Stuttgart · New York



Scheme 2 Flow synthesis of oxazole carboxylic acid 10

heated or cooled to a desired temperature. The reaction stream is then subjected to in-line purification using solidsupported quenching and scavenger resins prior to the product being collected or relayed directly into the next step of the synthesis sequence. Using this flow configuration we established conditions for converting dimethoxycinnamic acid (6) into oxazoline derivative 9 as shown in Scheme 2. The carboxylic acid was activated with CDI and coupled with threonine tert-butyl ester hydrochloride (7) in situ before adding a third stream containing diethylaminosulfur trifluoride (DAST)¹² to accomplish the cyclodehydration of the intermediate amide. A set of heterogeneous scavengers such as a sulfonic acid (QP-SA,¹³ to remove Et₃N, imidazole and residual threonine tert-butyl ester), a tertiary amine (QP-DMA,¹³ to remove residual acid), and plugs of CaCO₃ and SiO₂ (to quench and trap DAST and HF) were used in glass columns affording oxazoline 9 in greater than 95% purity. In a second step this oxazoline was then oxidized to the corresponding oxazole using a mixture of BrCCl₃ and DBU¹⁴ followed by cleavage of the *tert*-butyl ester using the previous mentioned QP-SA scavenger resin placed in a heated glass column.

Using this flow sequence we achieved rapid access to pure carboxylic acid **10** as a white, bench-stable solid in multigram quantities without the need to perform any traditional workup or purification steps. This material could then be quantitatively converted into acyl chloride **3** using oxalyl chloride in the presence of catalytic amounts of DMF by a standard batch method.

Having accomplished an efficient synthesis of the lefthand fragment 10, we next focused on the preparation of a suitable coupling partner. As the central carbon-carbon bond C5–C6 can be formed by a formal acylation, we synthesized several acetyl oxazoles 4a-c. Whilst oxazole nitrile 4a is a known compound,¹⁵ oxazole diester building blocks 4b and 4c were prepared in two steps from diazo esters 11 and 12¹⁶ as described in Scheme 3. An NH insertion of the rhodium carbenoid derived from 11 and 12 with methyl malonate monoamide¹⁷ led to keto amides **13** and 14, which cyclized to oxazoles 4b and 4c under Wipf's conditions.¹⁸ As this sequence involved the slow addition of methyl malonate monoamide, the use of a standard syringe pump gave the best results. This was followed by cyclodehydration of intermediates 13 and 14 where again all steps were performed by convenient batch methods (Scheme 3).

In order to progress the synthesis, acid chloride **3** was coupled by a Claisen condensation with the oxazole esters **4a**–**c** (Scheme 4). It was observed that the diesters **4b** and **4c** were prone to self-condensation upon treatment with NaHMDS, therefore the base (2.3 equiv) was added, via a syringe pump to the mixture of acid chloride **3** (1.3 equiv) and oxazole esters **4a**–**c** in THF (1.0 equiv each) at –78



Scheme 3 Preparation of oxazoles 4b and 4c. *Reagents and conditions*: (a) methyl malonate monoamide, $Rh_2(OAc)_4$ (2 mol%), CH_2Cl_2 , reflux, syringe pump addition, 84% for 13, 56% for 14; (b) I₂, Ph₃P, Et₃N, CH_2Cl_2 , r.t., 55%; (c) I₂, Ph₃P, *i*-Pr₂NEt, PhMe, 80 °C, 71%.



Scheme 4 Claisen condensation of acid chloride 3 with oxazoles 4a–c. *Reagents and conditions*: (a) NaHMDS (2.3 equiv) added at –78 °C, via a syringe pump, to 3 (1.3 equiv) and 4a–c (1.0 equiv) in THF.

°C. This procedure cleanly generated the desired adducts **15a–c**, which were employed without purification in the subsequent decarboxylation step.

The use of a polymer-supported base for the Claisen condensation of acid chloride **3** with oxazole esters **4a–c** was also examined. As illustrated in Scheme 5, stock solutions of **3** and **4a** in MeCN were mixed in a microreactor, followed by elution through a cartridge¹⁹ containing PS-BE-MP.²⁰ Unfortunately this procedure did not deliver the expected coupled material. Ultimately, it was found that both the product and excess starting material **4a** were retained as the corresponding enolates by the immobilized base. This was potentially very problematic because applying any standard release protocol involving displacement of the product from the column by treatment with an acidic solution would also liberate the excess starting material and acid via hydrolysis of the acyl chloride **3**. As a result additional processing through subsequent in-line scavenging or column chromatography of the crude product would be required. However, pleasingly it was discovered that when LiBr was added to the stock solution of ester **4a**, the desired product **15a** could then be obtained cleanly in 85% yield after aqueous quenching. It is proposed that the addition of the LiBr leads to the formation of the strongly chelated and highly soluble lithium enolate adduct **16**. This therefore enables flow processing of the reaction with the advantage that the major potential contaminants of excess **4a** and hydrolyzed carboxylic acid of **3** are more strongly bound on the polymeric support and so removed from the product stream of enolate **16**.

Next the decarboxylation of Claisen adducts **15a–c** was studied in detail (Table 1). Adduct **15a** cleanly underwent decarboxylation in the presence of excess NaCl (20 equiv) in wet DMSO, to produce bisoxazole nitrile **17a** in 73% overall yield (two steps from **4a**). When the corresponding bismethylester **15b** was reacted under identical condi-



Scheme 5 Claisen reaction of oxazole 3 and 4a induced by PS-BEMP as polymer-supported base in the presence of LiBr

Synlett 2011, No. 10, 1375-1380 © Thieme Stuttgart · New York

tions, surprisingly, no easily identified products could be obtained, and recovery of the starting material after chromatographic separation was poor. However, decarboxylation of **15b** in the absence of NaCl did allow for the isolation of product **17b** albeit in very low yield (7%). The salt-free decarboxylation of adduct **15c**, bearing a phenyl ester group at C-2 (instead of the methyl ester) on the other hand produced bisoxazole ester **17c** in a correspondingly higher 40% overall yield from **4c**.

These results suggest that in the case of substrate 15b, with a methyl ester in the 2-position, both the B_{AL}^2 and B_{AC}^{2} cleavage mechanisms operate with almost the same efficiency. This is supported by the observation that although both pathways might be possible in the presence of chloride ions, presumably only the latter mechanism is possible when water is the sole nucleophile.²¹ However, the disappointingly low yield of bisoxazole 17b (7%) from the salt-free reaction indicates that the C-2 methyl ester is highly reactive towards hydrolysis at elevated temperatures, and so it was not possible to tune the reactivity by switching the external nucleophile from chloride to water. Furthermore, it would appear that the free C-2 carboxylate generated from 15b is highly unstable and prone to thermal decarboxylation, followed by a further decomposition of the resulting C-2 carbanion precluding this as a viable synthetic route. Compared to substrate 15b, the alternative diester substrate 15c demonstrated improved stability, as the BAL2 mechanism cannot operate at the site of the C-2 phenyl ester. Thus, it was possible to isolate the decarboxylation product **17c** in 40% yield. This was lower than the 73% yield of the C-2 cyano compound 17a, due to partial hydrolysis of the phenyl ester of 15c via the B_{AC}^{2} mechanism.

Table 1Decarboxylation of Keto Esters 15a-c to BisoxazoleKetones 15a-c



Nevertheless, the bisoxazole nitrile **17a** was not a suitable precursor to the natural product **2** as the cyano group at C-2 resisted all attempts at hydrolysis to the carboxylic acid. Even strongly acidic or basic conditions only led to the

Synlett 2011, No. 10, 1375–1380 © Thieme Stuttgart · New York

corresponding carboxamide in low yields, accompanied by significant demethylation of the C-20 methoxy group. Pleasingly, however, bisoxazole phenyl ester **17c** readily hydrolyzed when treated with barium hydroxide generating the corresponding acid, which was obtained in 1:1 mixture with phenol. Purification and isolation of the acid was possible, however, it was found that the crude material could be coupled with pentadienyl amine **5**²² directly even in the presence of the phenol (Scheme 6). When the acid was activated with TBTU²³ (1.5 equiv), and amine **5** (2.2 equiv) was added to the mixture after 15 minutes incubation, no product derived from phenol addition was observed, with *O*-methyl siphonazole (**2**)²⁴ being formed exclusively, and enabling isolation in 47% combined yield over the two steps.



Scheme 6 Conversion of bisoxazole ester **17c** into *O*-methyl siphonazole **2**. *Reagents and conditions*: (a) BaOH₂·8H₂O, THF–H₂O, r.t.; (b) TBTU (1.5 equiv), **5** (2.2 equiv), DMF, r.t., 47% (2 steps).

The outcome of our efforts presented in this article shows that the synthesis of the natural product O-methyl siphonazole (2) from oxazole units 3 and 4c, and amine 5 was accomplished via a short sequence including a Claisen condensation and Krapcho decarboxylation as key steps. Furthermore, the use of flow techniques in concert with immobilized reagents and scavengers greatly improved the overall efficiency of the synthesis by circumventing unnecessary isolation and purification steps. In summary, this work exemplifies the benefit of an integrated approach combining traditional batch procedures with appropriate enabling techniques such as flow chemistry. This strategy afforded O-methyl siphonazole in nine steps.

Acknowledgment

The authors thank the Deutsche Forschungsgemeinschaft (M. Brasholz), the Cambridge European Trust, and the Ralph Raphael Studentship (M. Baumann), the Royal Society (I.R.B.), the EPSRC (N.N. and J.J.H.) and the BP endowment (S.V.L.) for financial support.

References and Notes

- (a) Bull, J. A.; Balskus, E. P.; Horan, R. A. J.; Langner, M.; Ley, S. V. Angew. Chem. Int. Ed. 2006, 45, 6714. (b) Bull, J. A.; Balskus, E. P.; Horan, R. A. J.; Langner, M.; Ley, S. V. Chem. Eur. J. 2007, 13, 5515. (c) Enriquez-Garcia, A.; Ley, S. V. Collect. Czech. Chem. Commun. 2009, 74, 887.
- (2) Nett, M.; Erol, ; Kehraus, S.; Köck, M.; Krick, A.; Eguevera, E.; Neu, E.; König, G. M. Angew. Chem. Int. Ed. 2006, 45, 3863.

- (3) (a) Linder, J.; Moody, C. J. *Chem. Commun.* 2007, 1508.
 (b) Linder, J.; Blake, A. J.; Moody, C. J. *Org. Biomol. Chem.* 2008, *6*, 3908.
- (4) (a) Zhang, J.; Ciufolini, M. A. Org. Lett. 2009, 11, 2389.
 (b) Zhang, J.; Polishchuk, E. A.; Chen, J.; Ciufolini, M. A. J. Org. Chem. 2009, 74, 9140.
- (5) (a) Noël, T.; Naber, J. R.; Hartman, R. L.; McMullen, J. P.; Jensen, K. F.; Buchwald, S. L. Chem. Sci. 2011, 2, 287. (b) Ceylan, S.; Coutable, L.; Wegner, J.; Kirschning, A. Chem. Eur. J. 2011, 17, 1884. (c) Polyzos, T.; O'Brien, M.; Petersen, T. P.; Baxendale, I. R.; Ley, S. V. Angew. Chem. Int. Ed. 2011, 50, 1190. (d) Ngamsom, B.; Hickey, A. M.; Greenway, G. M.; Littlechild, J. A.; McCreedy, T.; Watts, P.; Wiles, C. Org. Biomol. Chem. 2010, 8, 2419. (e) Bagley, M. C.; Fusillo, V.; Jenkins, R. L.; Lubinu, M. C.; Mason, C. Org. Biomol. Chem. 2010, 8, 2245. (f) Wegner, J.; Ceylan, S.; Friese, C.; Kirschning, A. Eur. J. Org. Chem. 2010, 4372. (g) McMullen, J. P.; Stone, M. T.; Buchwald, S. L.; Jensen, K. F. Angew. Chem. Int. Ed. 2010, 49, 7076. (h) McMullen, J. P.; Jensen, K. F. Org. Process Res. Dev. 2010, 14, 1169. (i) Damm, M.; Glasnov, T. N.; Kappe, C. O. Org. Process Res. Dev. 2010, 14, 215. (j) Riva, E.; Gagliardi, S.; Martinelli, M.; Passarella, D.; Vigo, D.; Rencurosi, A. Tetrahedron 2010, 66, 3242. (k) Amemiya, F.; Fuse, K.; Fuchigami, T.; Atobe, M. Chem. Commun. 2010, 46, 2730. (1) Brandt, J. C.; Elmore, S. C.; Robinson, R. I.; Wirth, T. Synlett 2010, 3099. (m) Nagaki, A.; Takabayasi, N.; Tomida, Y.; Yoshida, J. Beilstein J. Org. Chem. 2009, 5, No. 16. (n) Gustafsson, T.; Pontén, F.; Seeberger, P. H. Chem. Commun. 2008, 26, 1100. (o) Sahoo, H. R.; Kralji, J. G.; Jensen, K. F. Angew. Chem. Int. Ed. 2007, 46, 5704. (p) Mason, B. P.; Price, K. E.; Steinbacher, J. L.; Bogdan, A. R.; MacQuade, D. T. Chem. Rev. 2007, 107, 2300.
- (6) (a) Baumann, M.; Baxendale, I. R.; Ley, S. V. *Mol. Diversity* 2011, in press; DOI: 10.1007/s11030-010-9282-1.
 (b) Bogdan, A. R.; James, K. *Chem. Eur. J.* 2010, *16*, 14506.
 (c) Hessel, V. *Chem. Eng. Technol.* 2009, *32*, 1655.
 (d) Ley, S. V.; Baxendale, I. R. *Chimia* 2008, *62*, 162.
- (7) (a) Irfan, M.; Glasnov, T. N.; Kappe, C. O. Org. Lett. 2011, 13, 984. (b) Martin, L. J.; Marzinzik, A. L.; Ley, S. V.; Baxendale, I. R. Org. Lett. 2011, 13, 320. (c) Smith, C. J.; Nikbin, N.; Ley, S. V.; Lange, H.; Baxendale, I. R. Org. Biomol. Chem. 2011, 9, 1938. (d) Malet-Sanz, L.; Madrzak, J.; Ley, S. V.; Baxendale, I. R. Org. Biomol. Chem. 2010, 8, 5324. (e) McPake, C. B.; Murray, C. B.; Sandford, G. Chimica oggi 2010, 26, 6. (f) O'Brien, M.; Baxendale, I. R.; Ley, S. V. Org. Lett. 2010, 12, 1596. (g) Bartrum, H. E.; Blakemore, D. C.; Moody, C. J.; Hayes, C. J. J. Org. Chem. 2010, 75, 8674. (h) Brandt, J. C.; Wirth, T. Beilstein J. Org. Chem. 2009, 5, No. 30. (i) Bogdan, A. R.; Sach, N. W. Adv. Synth. Catal. 2009, 351, 849. (j) Baumann, M.; Baxendale, I. R.; Nikbin, N.; Ley, S. V.; Smith, C. D.; Tierney, J. P. Org. Biomol. Chem. 2008, 6, 1577. (k) Wiles, C.; Watts, P. Eur. J. Org. Chem. 2008, 5597. (1) Sandford, G. J. Fluorine Chem. 2007, 128, 90.
- (8) (a) Hodgkinson, J. T.; Galloway, W. R. J. D.; Shreya, S.; Baxendale, I. R.; Ley, S. V.; Ladlow, M.; Welch, M.; Spring, D. R. Org. Biomol. Chem. 2011, 9, 57. (b) Carter, C. F.; Baxendale, I. R.; Pavey, J. B. J.; Ley, S. V. Org. Biomol. Chem. 2010, 8, 1588. (c) Wheeler, R. C.; Benali, O.; Deal, M.; Farrant, E.; MacDonald, S. J. F.; Warrington, B. H. Org. Process Res. Dev. 2007, 11, 704. (d) Hornung, C. H.; Mackley, M. R.; Baxendale, I. R.; Ley, S. V. Org. Process Res. Dev. 2007, 11, 399. (e) Smith, C. J.; Iglesias-Sigüenza, F. J.; Baxendale, I. R.; Ley, S. V. Org. Biomol. Chem. 2007, 5, 2758. (f) Roberge, D. M.; Ducry, L.; Bieler, N.; Cretton,

P.; Zimmermann, B. Chem. Eng. Technol. 2005, 28, 318.

- (9) (a) Rasheed, M.; Wirth, T. Angew. Chem. Int. Ed. 2011, 50, 357. (b) Smith, C. J.; Smith, C. D.; Nikbin, N.; Ley, S. V.; Baxendale, I. R. Org. Biomol. Chem. 2011, 9, 1927.
 (c) Venturoni, F.; Nikbin, N.; Ley, S. V.; Baxendale, I. R. Org. Biomol. Chem. 2010, 8, 1798. (d) Baxendale, I. R.; Griffiths-Jones, C. M.; Ley, S. V.; Tranmer, G. K. Synlett 2006, 427.
- (10) (a) Baumann, M.; Baxendale, I. R.; Kirschning, A.; Ley, S. V.; Wegner, J. Heterocycles 2011, 82, 1297. (b) Hopkin, M. D.; Baxendale, I. R.; Ley, S. V. Chem. Commun. 2010, 46, 2450. (c) Qian, Z.; Baxendale, I. R.; Ley, S. V. Chem. Eur. J. 2010, 16, 12342. (d) Brasholz, M.; Johnson, B. A.; Macdonald, J. M.; Polyzos, A.; Tsanaktsidis, J.; Saubern, S.; Holmes, A. B.; Ryan, J. H. Tetrahedron 2010, 66, 6445. (e) Tamborini, L.; Conti, P.; Pinto, A.; Micheli, C. D. Tetrahedron: Asymmetry 2010, 21, 222. (f) Herath, A.; Dahl, R.; Cosford, N. D. P. Org. Lett. 2010, 12, 412. (g) Baxendale, I. R.; Schou, S.; Sedelmeier, J.; Ley, S. V. Chem. Eur. J. 2010, 16, 89. (h) Baxendale, I. R.; Ley, S. V.; Mansfield, A. C.; Smith, C. D. Angew. Chem. Int. Ed. 2009, 48, 4017. (i) Baxendale, I. R.; Deeley, C. M.; Griffiths-Jones, C. M.; Ley, S. V.; Saaby, S.; Tranmer, G. K. Chem. Commun. 2006, 24, 2566. (j) Baxendale, I. R.; Ley, S. V.; Smith, C. D.; Tranmer, G. K. Chem. Commun. 2006, 4835.
- (11) Vapourtec[©] R2+/R4 system was used; website: www.vapourtec.co.uk.
- (12) (a) Middleton, W. J. J. Org. Chem. 1975, 40, 574.
 (b) Phillips, A. J.; Uto, Y.; Wipf, P.; Reno, M. J.; Williams, D. R. Org. Lett. 2000, 2, 1165. Use of DAST in flow:
 (c) Baumann, M.; Baxendale, I. R.; Ley, S. V. Synlett 2008, 2111. (d) Baumann, M.; Baxendale, I. R.; Martin, L. J.; Ley, S. V. Tetrahedron 2009, 65, 6611.
- (13) QP-SA and QP-DMA resins are commercially available from Johnson Matthey. Website: http://www.matthey.com/.
- (14) Williams, D. R.; Lowder, P. D.; Gu, Y.-G.; Brooks, D. A. *Tetrahedron Lett.* **1997**, *38*, 331.
- (15) Doleschall, G.; Seres, P. J. Chem. Soc., Perkin Trans. 1 1988, 1875.
- (16) Compound 12 was prepared by ring opening of 2,2,6-trimethyl-4*H*-1,3-dioxin-4-one with phenol, see: Clemens, R. J.; Hyatt, J. A. J. Org. Chem. 1985, 50, 2431. Followed by diazo group transfer, see: Tullis, J. S.; Helquist, P. Org. Synth. 1997, 74, 229.
- (17) Bagley, M. C.; Buck, R. T.; Hind, S. L.; Moody, C. J. J. Chem. Soc., Perkin Trans. 1 1998, 591.
- (18) Wipf, P.; Miller, C. P. J. Org. Chem. 1993, 58, 3604.
- (19) An Omnifit column was used, www.omnifit.com.
- (20) For related applications of PS-BEMP iminophosphorane base, see: (a) Bensa, D.; Constantieux, T.; Rodriguez, J. *Synthesis* 2004, 923. (b) Baumann, M.; Baxendale, I. R.; Ley, S. V.; Smith, C. D.; Tranmer, G. K. *Org. Lett.* 2006, *8*, 5231.
- (21) (a) Krapcho, A. P.; Jahngen, E. G. E. Jr.; Lovey, A. J. *Tetrahedron Lett.* **1974**, *15*, 1091. (b) Krapcho, A. P.; Weimaster, J. F.; Eldridge, J. M.; Jahngen, E. G. E. Jr.; Lovey, A. J.; Stephens, W. P. *J. Org. Chem.* **1978**, *43*, 138.
- (22) (a) Mori, K. *Tetrahedron* 1974, *30*, 3810. (b) Grieco, P. A.;
 Galatsis, P.; Spohn, R. F. *Tetrahedron* 1986, *42*, 2847.
- (23) Poulain, R. F.; Tartar, A. L.; Déprez, B. P. Tetrahedron Lett. 2001, 42, 1495.
- (24) **NMR Data** ¹H NMR (600 MHz, CDCl₃): $\delta = 2.64, 2.67$ (2 s, 2 × 3 H, 18-H, 19-H), 3.92, 3.93 (2 s, 2 × 3 H, 20-H, 21-H), 4.03 (*t*, J = 5.8 Hz, 2 H, 1'-H), 4.41 (s, 2 H, 5-H), 5.06 (d, J = 10.3Hz, 1 H, 5'-H), 5.18 (d, J = 17.0 Hz, 1 H, 5'-H), 5.73 (td, J = 5.8, 15.0 Hz, 1 H, 2'-H), 6.22 (dd, J = 10.3, 15.0 Hz, 1 H,

Synlett 2011, No. 10, 1375–1380 © Thieme Stuttgart · New York

3'-H), 6.31 (td, J = 10.3, 17.0 Hz, 1 H, 4'-H), 6.75 (d, J = 16.4, 1 H, 10-H), 6.88 (d, J = 8.2 Hz, 1 H, 16-H), 7.00 (t, J = 5.8 Hz, 1 H, NH), 7.07 (d, J = 1.6 Hz, 1 H, 13-H), 7.09 (dd, J = 1.6, 8.2 Hz, 1 H, 17-H), 7.44 (d, J = 16.4 Hz, 1 H, 11-H) ppm. ¹³C NMR (150 MHz): $\delta = 11.7$, 12.4 (2 q, C-18, C-19), 39.4 (t, C-5), 40.3 (t, C-1'), 55.9, 56.0 (2 q, C-20, C- 21), 108.9 (d, C-13), 110.8 (d, C-10), 111.2 (d, C-16), 117.4 (t, C-5'), 121.5 (d, C-17), 128.1 (s, C-12), 129.45, 129.49 (2 d, C-2', C-3'), 132.8 (s, C-2), 134.5 (s, C-7), 136.1 (d, C-4'), 137.3 (d, C-11), 149.3, 150.6 (2 s, C-14, C-15), 153.8 (s, C-3), 155.3 (s, C-8), 155.6 (s, C-4), 159.1 (s, C-9), 161.7 (s, C-1), 189.4 (s, C-6) ppm.