

# Synthesis of the Alkaloid Natural Products (+)-Plicane and (–)-Obliquine, Using Polymer-Supported Reagents and Scavengers<sup>†</sup>

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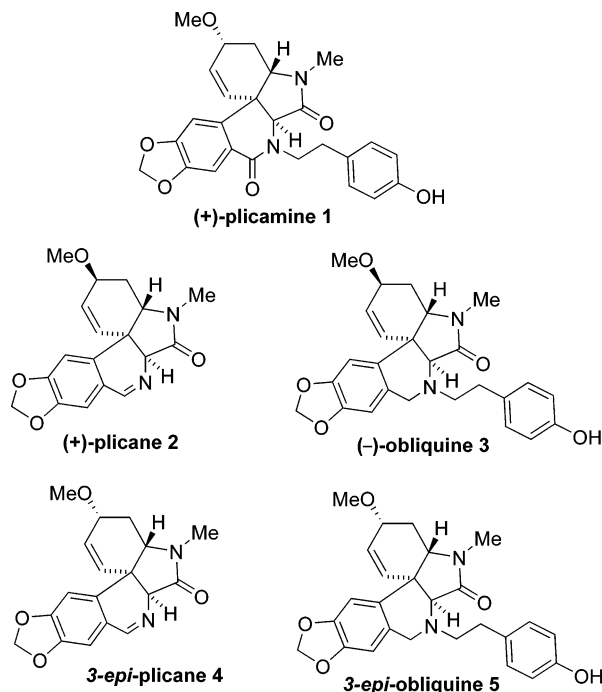
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Two new naturally occurring amaryllidaceae alkaloids have been synthesized, using a divergent approach facilitated by the use of polymer-supported reagents and scavengers.

## Introduction

The amaryllidaceae alkaloids have been the recipients of considerable synthetic interest, because of their extensive structural diversity and broad biological activity.<sup>1,2</sup> Our group has already reported on the synthesis of a selection of these compounds that have been prepared using a suite of solid-supported reagents,<sup>3–5</sup> to facilitate more-efficient synthesis.<sup>6</sup> We were particularly interested in the molecular architecture of (+)-plicamine (**1**) (Figure 1), which is the first member of a new *bis*-nitrogen-containing family of alkaloids to be isolated<sup>5</sup> from extracts of the Turkish *Galanthus plicatus* (subsp. *Byzantinus*).<sup>8</sup> Following our synthesis of this molecule,<sup>6b,c</sup> we became aware of a subsequent study by the same group in which a structurally related alkaloid (+)-plicane (**2**) was characterized.<sup>9</sup> In addition, an investigation into a related amaryllidaceae *Cyrtanthus obliquus*,<sup>10</sup> which is indigenous to the Cape and KwaZulu Natal provinces of South Africa, provided a third member of the series, namely, (–)-obliquine (**3**).<sup>11</sup> It would appear from the structural similarities that the compounds of the plicamine subgroup can exhibit both configurations at C-3. We anticipated that the synthetic pathway in our original synthesis of plicamine<sup>6b,c</sup> would allow us to access these two new natural products and also permit the preparation of their two epimeric derivatives (**4** and **5**) in a rapid and divergent fashion. This would also assist structural assignment, because ambiguity could easily exist in the series and in addition to providing new molecules for biological screening.

Therefore, starting from the common intermediate **6**,<sup>6b,c</sup> two parallel pathways were followed (Scheme 1). Route A involved the initial conversion of the C-3 alcohol to the resulting mesylate, followed by stereocontrolled nucleophilic inversion with methanol to yield **7** in high yield. Alternatively, route B furnished the C-3 methoxy epimer **8** via methylation using trimethylsilyl diazomethane (TMS–CHN<sub>2</sub>) and an immobilized sulfonic acid catalyst also in excellent yield. The trifluoroacetate groups from both compounds **7** and **8** could be efficiently cleaved under basic conditions when facilitated by flash microwave heating<sup>12</sup> to give the corresponding amines **9** and **10**, respectively. Direct oxidation of amine **9** with cerium ammonium nitrate absorbed on silica then gave a clean transformation to the imine, (+)-plicane (**2**). In an identical manner, 3-*epi*-plicane (**4**) could be prepared from the matching amine **10**, using the same reagent.



**Figure 1.** Schematic of the plicamine family of dinitrogenous amaryllidaceae alkaloids **1–5**.

Branching in a different direction from the same two amines **9** and **10** permitted synthesis of the products (–)-obliquine (**3**) and *epi*-obliquine (**5**) (subsequently leading to plicamine (**1**)). *N*-Alkylation with the alkyl bromide **11** promoted by an immobilized carbonate base, followed by scavenging the reaction mixture with an aminothiols resin, proved to be an efficient sequence. The natural products were identical to the reported authentic material.

**Conclusion.** In conclusion, a general and straightforward synthesis design has led to the preparation of a range of naturally occurring amaryllidaceae alkaloids. In addition, we have prepared two structural analogues (**4** and **5**) that possess the required functional components and stereochemistry of products that are likely to be isolated members of the plicamine family in the future. The synthesis strategy that is applied relies on the application of solid-supported reagents to provide products and intermediates in high yield and purity, without the need to resort to extensive chromatographic purification following each synthesis step.

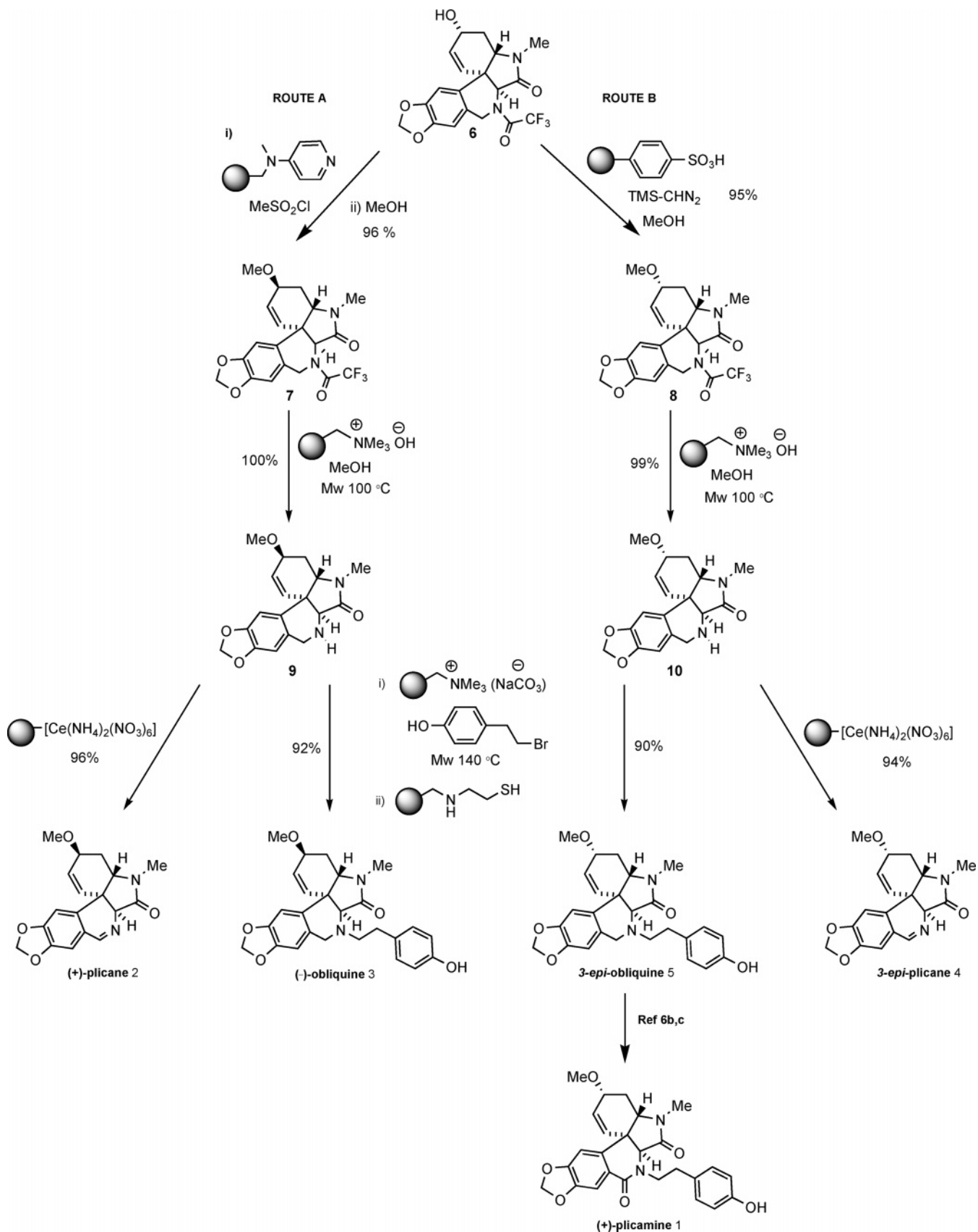
## Experimental Section

<sup>1</sup>H NMR spectra were recorded on a Bruker Advance DPX-400 spectrometer, with residual chloroform as the

<sup>†</sup> This paper is dedicated to Dave Sherrington, a good friend and fine scientist on the occasion of his 60th birthday.

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Scheme 1. Synthesis Scheme for the Preparation of Plicamine (1) and Its Derivatives



internal reference ( $\delta_{\text{H}} = 7.26$  ppm).  $^{13}\text{C}$  NMR spectra were recorded in  $\text{CDCl}_3$  on the same spectrometers, with the central peak of chloroform as the internal reference ( $\delta_{\text{C}} = 77.0$  ppm). DEPT 135 was used to aid in the assignment of signals in the  $^{13}\text{C}$  NMR spectra.

**Plicane (2).** Rf 2.530, MS 327.1 ( $\text{MH}^+$ ).  $[\alpha]_{\text{D}} = 285.9$  ( $c = 0.15$  in  $\text{MeOH}$ ); IR(neat),  $\nu_{\text{max}}$ : 3450, 2932, 2850, 1696, 1644, 1609, 1599, 1488, 1386, 1315, 1260, 1186, 1167, 1090, 1028, 987, 837, 820, 815  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta = 8.20$  (1 H, s, H-8), 6.85 (1 H, s, H-12),

6.78 (1 H, s, H-9), 6.07 (1 H, dd,  $J = 10.15, 0.85$  Hz, H-2), 5.94 (2 H, s, OCH<sub>2</sub>O), 5.42 (1 H, d,  $J = 10.15$  Hz, H-1), 4.22 (1 H, br. s, H-6a), 3.73 (1 H, m, H-3), 3.60 (1 H, m, H-4a), 3.39 (3 H, s, OMe), 2.81 (3 H, s, NMe), 2.11 (1 H, m, H-4<sub>B</sub>), 1.88 (1 H, m, H-4<sub>A</sub>). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 172.4$  (C-6), 158.4 (C-8), 151.0 (C-11), 147.1 (C-10), 131.3 (C-12a), 130.6 (C-1), 129.4 (C-2), 123.1 (C-8<sub>A</sub>), 109.2 (C-9), 106.9 (C-12), 101.3 (OCH<sub>2</sub>O), 71.5 (C-3), 66.6 (C-6a), 62.3 (C-4a), 56.6 (OMe), 43.6 (C-12b), 30.6 (NMe), 28.4 (C-4). HR-MS Calcd for C<sub>18</sub>H<sub>18</sub>N<sub>2</sub>O<sub>4</sub>-Na: 349.1164. Found: 349.1170.

**Obliquine (3).** Rf 2.384, MS 449.1 (MH<sup>+</sup>). [ $\alpha$ ]<sub>D</sub> = -137.8 ( $c = 0.35$  in MeOH); IR(neat),  $\nu_{\max}$ : 3311.5, 2970.2, 1716.1, 1670.2, 1513.3, 1488.1, 1456.1, 1400.0, 1233.6, 1158.4, 1108.0, 1036.2, 944.7, 935.7, 933.1 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.01$  (2 H, d,  $J = 8.5$  Hz, H-11'), 6.73 (2 H, d,  $J = 8.5$  Hz, H-10'), 6.55 (1 H, s, H-12), 6.46 (1 H, s, H-9), 5.96 (1 H, dd,  $J = 10.0, 5.2$  Hz, H-2), 5.90 (1 H, d,  $J = 10.0$  Hz, H-1), 5.87 (2 H, s, OCH<sub>2</sub>O), 3.98 (1 H, d,  $J = 15.25$  Hz, H-8<sub>A</sub>), 3.89 (1 H, m, H-3), 3.66 (1 H, dd,  $J = 11.5$  and 4.5 H-4<sub>A</sub>), 3.70 (1 H, d,  $J = 15.25$  Hz, H-8<sub>B</sub>), 3.66 (1 H, s, H-6<sub>A</sub>), 3.48 (3 H, OMe), 3.31 (1 H, m, H-8<sub>A</sub>'), 3.27 (1 H, m, H-8<sub>B</sub>'), 2.75 (2 H, t,  $J = 7.9$  Hz, H-7'), 2.79 (3 H, s, NMe), 2.26 (1 H, m, H-4<sub>B</sub>), 1.89 (1 H, m, H-4<sub>B</sub>). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta = 171.3$  (C-6), 154.8 (C-4'), 147.1, 147.0 (C-10 and C-11), 135.5 (C-1), 132.8 (C-9'), 131.5 (C-12a), 129.5 (C-8a), 129.8 (C-10'), 124.8 (C-2), 115.4 (C-11'), 109.0 (C-9), 107.8 (C-12), 101.9 (OCH<sub>2</sub>O), 72.5 (C-3), 66.5 (C-6a), 62.6 (C-4a), 56.8 (C-8'), 56.5 (OMe), 50.6 (C-8), 44.7 (C-12b), 33.9 (C-7'), 32.1 (C-4), 27.8 (NMe). HR-MS Calcd for C<sub>26</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>-Na: 471.1896. Found: 471.1892.

**3-Epi-plicane (4).** Rf 2.426, MS 327.1 (MH<sup>+</sup>). [ $\alpha$ ]<sub>D</sub> = 88.7 ( $c = 0.15$  in MeOH); IR(neat),  $\nu_{\max}$ : 3288, 2935, 1675, 1644, 1605, 1525, 1488, 1399, 1234, 1140, 950, 945, 820 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.23$  (1 H, s, H-8), 6.81 (1 H, s, H-12), 6.62 (1 H, s, H-9), 6.04 (1 H, dd,  $J = 10.2, 1.35$  Hz, H-2), 5.96 (2 H, s, OCH<sub>2</sub>O), 5.50 (1 H, d,  $J = 10.2$  Hz, H-1), 4.39 (1 H, br. s, H-6a), 3.84 (1 H, m, H-3), 3.56 (1 H, dd,  $J = 12.5, 4.8$  Hz, H-4<sub>A</sub>), 3.34 (3 H, s, OMe), 2.80 (3 H, s, NMe), 2.13 (1 H, m, H-4<sub>B</sub>), 1.86 (1 H, m, H-4<sub>A</sub>). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 171.5$  (C-6), 159.4 (C-8), 151.9 (C-11), 147.0 (C-10), 131.8 (C-12a), 130.9 (C-1), 130.3 (C-2), 120.9 (C-8a), 109.9 (C-9), 108.3 (C-12), 101.3 (OCH<sub>2</sub>O), 73.1 (C-3), 67.1 (C-6a), 62.5 (C-4a), 56.5 (OMe), 40.6 (C-12b), 30.1 (NMe), 28.3 (C-4). X-ray crystal structure obtained of the racemic material registered with the Cambridge Crystallographic Database, under reference CDS 252364.

**3-Epi-obliquine (5).** Rf 2.721, MS 449.1 (MH<sup>+</sup>). [ $\alpha$ ]<sub>D</sub> = 143.6 ( $c = 0.25$  in MeOH); IR(neat),  $\nu_{\max}$ : 3293, 2927, 1669, 1613, 1515, 1483, 1454, 1386, 1234, 1102, 1036, 932, 830 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.00$  (2 H, d,  $J = 8.3$  Hz, H-11'), 6.75 (2 H, d,  $J = 8.3$  Hz, H-10'), 6.50 (1 H, d, H-Ar), 6.46 (1 H, d, H-Ar), 5.93 (1 H, d,  $J = 10.15$  Hz, H-1), 5.89 (2 H, s, CH<sub>2</sub>O<sub>2</sub>), 5.79 (1 H, d,  $J = 10.15$  Hz, H-2), 4.13 (1 H, m, H-3), 3.94 (1 H, m,  $J = 15.25$  Hz, H-8<sub>A</sub>), 3.78 (1 H, s, H-6a), 3.67 (1 H, d,  $J = 15.25$  Hz, H-8<sub>B</sub>), 3.56 (1 H, dd,  $J = 12.35, 4.35$  Hz, H-4a), 3.45 (3 H, s, OMe), 3.35 (1 H, m, H-8<sub>A</sub>'), 3.25 (1 H, m, H-8<sub>B</sub>'), 2.76 (5 H, m, H-7' and NMe), 2.53 (1 H, m, H-4<sub>A</sub>), 1.39 (1 H, m, H-4<sub>B</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 171.8$  (C-6), 154.7 (C-12'), 146.8 and 146.6 (C-10 and C-11), 133.1 (C-1), 128.3 (C-2), 132.5 (C-9'), 131.8 (C-12a), 129.6 (C-8a), 129.8 (C-10'), 115.4 (C-11'), 107.7 (C-9), 106.3 (C-12), 101.1 (C-15), 74.6 (C-

3), 66.4 (C-4a), 64.6 (C-6a), 56.9 (C-8'), 56.2 (OMe), 50.5 (C-8), 44.8 (C-12b), 34.0 and 32.0 (C-4 and C-7'), 27.9 (NMe). HR-MS Calcd for C<sub>26</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>-Na: 471.1896. Found: 471.1894.

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